Standard Operating Procedures: Cholera Toxin

Principal Investigator (print):	
Principal Investigator Signature:	
Date Reviewed:	
Location:	Rutgers University
Campus:	
Building:	
Designated Use Area / Room(s):	
Designated Storage Area/Room	
IBC Approval Number:	
IACUC Approval Number:	

Physical Characteristics:
Cholera toxin is the virulent factor from Vibrio cholera that is used in research for generally two purposes: it can be used as an activator of adenylate cyclase (subunit A), or it can be used as a tracer for neuronal connections (subunit B). One can purchase individual subunit-purified powders, or powder containing both subunits. Stock solutions of 1 – 2 mg/mL are made from the powder, then further diluted for use in both cell cultures and *in vivo*. It is highly toxic and can be fatal if swallowed or in contact with the skin. As such, proper handling is absolutely necessary.

Health Hazard Summary:

- The LD50 (lethal dose for 50%) in adult humans is 250 ug/kg
- It is highly toxic by ingestion and skin absorption as well as a general irritant. It can cause organ failure (bowels) and may be fatal.
- Inhalation May be harmful if inhaled. Causes respiratory tract irritation.
- Skin May be fatal if absorbed through skin. Causes skin irritation.
- Eyes Causes eye irritation.
- Ingestion May be fatal if swallowed.

Work must be conducted in either a ducted chemical hood or HEPA-filtered biosafety cabinet. Consult with REHS.

Safety Data Sheet (SDS): (Attach manufacturer-specific SDS to this SOP)
Read the manufacturer's SDS, formerly called the material safety data sheet (MSDS), and maintain a copy in your safety binder along with this SOP. For safety questions, contact Rutgers Environmental Health & Safety (REHS) at 848-445-2550.

Personnel Requirements:
All laboratory members working with cholera toxin must review the manufacturer specific SDS and be up to date on all required health and safety trainings.

Exposure Control:
- Purchase in quantities that can be dissolved at one time, within the original vial.
- Handle inside the chemical fume hood/certified biosafety cabinet (consult with REHS).
- Wear double nitrile gloves, eye protection and lab coat when handling.
- Cholera toxin must be kept in a secured location (e.g., locked freezer, box).
- Use syringe with integral safety feature, as applicable.
- Keep a solution of 10% bleach solution readily accessible (made fresh daily).
- Avoid inhalation and physical contact with.
- Ensure that a safety shower and eyewash station are nearby.

First Aid Procedures:
- *Call for medical advice immediately:*
  - Occupational Medicine Services (Newark) 973-972-2900
  - Hurtado Health Center (New Brunswick) 848-932-8254
  - Emergencies & After Hours – Call the Rutgers University Police Department (RUPD) or visit nearest hospital Emergency Room
    - 732-932-7211 (Piscataway & New Brunswick)
    - 973-972-4490 (RBHS - Newark / Scotch Plains)
    - 973-353-5111 (Rutgers-Newark)
- Additional first aid based on route of exposure:
  - *Ingestion/oral exposures* – rinse mouth with water.
  - *Inhalation exposure* – move person to fresh air and call for an ambulance if breathing becomes difficult.
  - *Contact exposure (eyes, nose, skin)* – flush the affected area with copious amounts of water for at least 15 minutes.
  - *Accidental Injection / Percutaneous* – call RUPD and request an ambulance or go to the nearest hospital emergency room.

Injury / Exposure Reporting:
Any exposure incidents must be reported in the REHS Accident Database located online at http://myrehs.rutgers.edu. The injured/exposed person's direct supervisor (e.g., PI or lab manager) needs to submit the incident report by the end of the work shift.

Spill Clean-up:

For small quantities (less than 5ml or 100 ug).

- If you don't feel comfortable cleaning up the spill, follow the instructions for large spills (below).
- Wear double nitrile gloves, lab coat, and safety glasses/goggles.
- Any broken glass fragments should be picked up with tongs, forceps or a small scoop (never use your fingers). Place the broken glass in a wide-mouthed plastic container. Tightly seal the container and contact REHS (http://rehs.rutgers.edu) for disposal.
- Liquids should be absorbed with paper towels and saturated with 10% bleach solution – 20 minute contact time!
- Solids should be wiped up with wetted paper towels saturated with a 10% bleach solution – 20 minute contact time!. Contaminated surfaces should then be cleaned three times using a detergent solution and paper towels followed by clean water.
- Inside a ducted hood, contaminated re-usable items (e.g., glassware and scoops) should be disinfected with a 10% bleach solution, washed three times with detergent by a trained employee wearing two pairs of nitrile gloves, eye protection and fully fastened lab coat or gown.
- Contaminated disposable items & spill clean-up waste (gloves, paper towels, absorbent pads, spill pads/pillows) must be bagged and autoclaved at 121°C and 15 psi for 60 minutes on liquid cycle (slow exhaust). The materials must then be disposed as biomedical waste.
- If your building does not have an autoclave, collect all spill clean-up materials in tightly sealed containers, and contact REHS (http://rehs.rutgers.edu) for disposal.

For large spills (greater than 5ml or 100 ug) or possible airborne :

- Evacuate the area.
- Report the spill to Rutgers University Police Department (RUPD)
    - 732-932-7211 (Piscataway & New Brunswick)
    - 973-972-4490 (RBHS - Newark / Scotch Plains)
    - 973-353-5111 (Rutgers-Newark)
- The police dispatcher will contact on-call REHS personnel.
- REHS staff will clean-up the spill.

General Safety Precautions for Dissolving and Aliquoting:

1. Post signs such as "Caution – Cholera Toxin" in the designated area(s) when working with the toxin until the cholera toxin has been returned to storage and the work area has been decontaminated.
2. Handle lyophilized powders and concentrated solutions in a fume hood, while wearing personal protective equipment (PPE): lab coat, safety glasses, and two pairs of nitrile gloves.
3. Place absorbent pad in the bottom of the hood/cabinet to contain potential spills.
4. Aliquot 5ml of a 10% bleach solution into a conical tube. Place the open tube in a tube rack to serve as waste receptacle for contaminated filtered pipette tips – minimum of 30 minute contact time!

5. Open vials carefully. If gloved finger touches the rim of the vial, change outer gloves immediately to avoid spreading contamination to other items.
6. Aliquot suspension into plastic tubes labeled: 'Cholera Toxin, concentration, your name and date'.
7. Place tubes in leak-proof secondary plastic container. Label container: 'Cholera Toxin', concentration, your name, date, TOXIN – DO NOT HANDLE'.
8. When ready for use, remove aliquot from freezer and let thaw to room temperature.
9. Use a pipette with filtered tip to transfer contents. Treat tips as described in #4 above.
10. Use extreme caution when preparing/handling needles of cholera toxin. Use needles with integral safety feature (e.g., BD Safety Glide™). Dispose of contaminated needles immediately in sharps container.
11. Animal Administration: restrain or anesthetize animal during the injection, label the cage card with a chemical hazard label and information, maintain cages in the ducted chemical hood, wear PPE when handling the animals, and collect carcasses of -administered animals in a separate bag with a chemical hazard warning label and cholera toxin information when returning carcasses to the research animal facility for disposal.
12. Animal Housing: use static or microisolator cages. Never use ventilated cage racks without first consulting with REHS.
13. Cages and Bedding: First bedding change (minimum 72-hour post-dosing):
    - Performed by laboratory personnel in biosafety cabinet.
    - Autoclave cages and bedding
    - Use biosafety cabinet to empty autoclaved bedding.
    - Dispose autoclaved bedding in biomedical waste container after first administration.
    - Subsequent changes performed by CMR staff. No special handling precautions.
14. Carcass Disposal: place in red biohazard bag and then into Vivarium biohazard freezer for incineration.
15. Inactivate cholera toxin stocks and -contaminated items by autoclaving (121^{o}C and 15 psi for 60 minutes) or chemical inactivation with sodium hypochlorite / bleach (30 minutes of contact time with liquid bleach) prior to disposal. Surfaces may be decontaminated with bleach.

Lab-Specific Procedures & Safety Precautions (to be completed by Principal Investigator). You may attach separate pages if more space is required.

Materials: List manufacturer, catalog number, quantity to be ordered and form of material – e.g., lyophilized powder.

Preparation: List specific steps for preparing aliquots, specify containment controls, PPE to be worn, disinfection steps for surfaces and equipment used, storage information.

Procedure for Use in Mice (if applicable): Include description of containment controls, injection dilution, method of injection, dosing and cage marking information, cage and bedding handling (including any restrictions for CMR/LAS personnel), carcass disposal, etc.

Procedure for Use In-Vitro (if applicable): Specify containment controls used, describe preparation of cell culture and how cell culture is treated and disposed.

Signatures:
By signing below, I certify that I have read this SOP and attached material, that I understand the procedures for working with cholera toxin, that I understand the hazards associated with using cholera toxin, and that I will use the procedures described in this SOP to safely handle and use cholera toxin.

Name (typed)	Job Title	Signature

References:
Appendix I - Guidelines for Work with Toxins of Biological Origin from the "Biosafety in Microbiological and Biomedical Laboratories" (BMBL) 5th Edition